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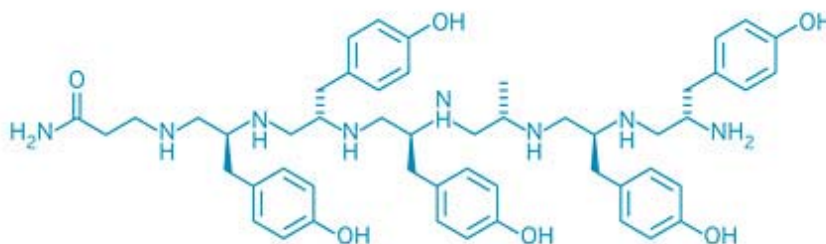
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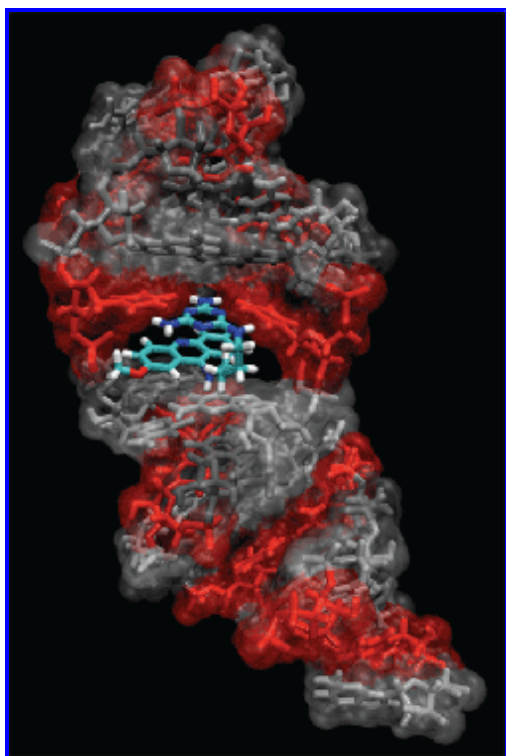
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## Targeting RNA

### Unique challenges face developers of drugs that hit disease-related RNAs rather than disease-related proteins

[Stu Borman](#)

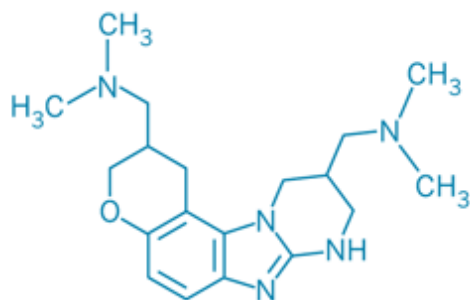
**HIV INHIBITOR** This multivalent-binding oligomer interferes with Tat-TAR binding, which boosts replication of the AIDS virus.



Courtesy of Anne Baranger and Steven Zimmerman

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IT'S A WRAP Model shows how triaminotriazine-acridine conjugate (cyan, red, blue, and white) may bind to mismatched U-U base pairs (red bands) in CUG-repeat mRNA (mostly gray).



### HCV ENEMY

Benzimidazoles like this one  
inhibit translation of  
hepatitis C virus genes.



Medical Center reported having exploited this advantage by developing a short string of nucleotides that recognizes and binds CUG-repeat mRNA and consequently reduces DM1 symptoms in mice ([C&EN, July 20](#), page 15; *Science* **2009**, 325, 336). Other groups are discovering nonnucleotide small molecules that can target DM1's mRNA. For example, bioorganic chemist [Benjamin L. Miller](#) of the University of Rochester Medical Center and coworkers recently used resin-bound dynamic combinatorial chemistry to identify potential DM1 drug leads. In this technique developed by Miller's group, high-affinity ligands from equilibrating combinatorial libraries are selected by competitive binding and then isolated by retention on a solid support.

With this approach, Miller and coworkers identified four peptide/small-molecule hybrids that bind CUG-repeat mRNA and inhibit its interaction with MBNL1 with low micromolar levels of activity (*J. Am. Chem. Soc.* **2008**, [130, 16254](#)). They are now trying to improve compound affinity and selectivity, in part by replacing the peptide components with peptidomimetics.

At the University of Illinois, Urbana-Champaign, chemistry professors [Anne M. Baranger](#) and [Steven C. Zimmerman](#) and coworkers are using rational drug design to find DM1 agents. Using structural information about CUG-repeat mRNA, they combined a sequence-selective binder, a nucleic acid-intercalating compound, and a linking group into a hybrid small molecule that they predicted would interact with the target (*Proc. Natl. Acad. Sci. USA*, DOI: [10.1073/pnas.0901824106](#)). The resulting triaminotriazine-acridine conjugate binds selectively to CUG-repeat mRNA and inhibits its ability to interact with MBNL1 with low micromolar activity. "We have shown that our compound is actually quite selective" for CUG-repeat mRNA, Zimmerman says.

**Also taking** the rational drug design route to small-molecule DM1 agents is assistant professor of chemistry [Matthew D. Disney](#) of the State University of New York, Buffalo, and coworkers. They compiled a database of RNA-binding ligands and the RNA structures or motifs to which the ligands bind. From that database, they identified the bisbenzimidazole Hoechst 33258 as a ligand for CUG-repeat mRNA. They then combined multiple copies to form a modularly assembled construct with improved affinity and specificity for the target (*J. Am. Chem. Soc.* **2009**, [131, 9767](#)).

A bisbenzimidazole pentamer they designed by this route inhibits with low nanomolar potency the formation not only of the CUG-repeat-mRNA/MBNL1 complex, but also of a repeat-mRNA/MBNL1 complex involved in spinocerebellar ataxia type 3, which causes lack of coordination of muscle movements. Disney and coworkers recently used the same database and modular assembly to find a lead compound for a closely related form of myotonic dystrophy (*ACS Chem. Biol.* **2009**, [4, 345](#)).

Researchers are also trying to find RNA-targeted medications for viral infections. For instance, at the University of California, San Diego, a team is studying how small molecules target the RNA genome of hepatitis C virus (HCV). HCV infection causes chronic hepatitis and hepatocellular carcinoma. New medications are urgently needed because current ones are weak and have serious side effects.

Benzimidazole derivatives were already known to bind to a subdomain of the internal ribosome entry site (IRES), the part of the HCV genome that causes host cells to translate viral RNA into proteins. Using fluorescence resonance energy transfer (FRET) and an IRES crystal structure they obtained earlier, the UC San Diego team determined the mechanism by which the benzimidazoles inhibit translation (*Nat. Chem. Biol.*, DOI: [10.1038/nchembio.217](#)).

Assistant professor of chemistry and biochemistry [Thomas Hermann](#) and coworkers carried out the work. They found that binding of one of the benzimidazoles causes a conformational change in the subdomain's hairpin-loop structure that prevents the IRES from interacting correctly with the ribosome. This, in turn, shuts off IRES-driven translation of viral genes in HCV-infected cells, short-circuiting the virus's life cycle.

Similar mechanisms have been found before for ribosome-targeting antibiotics, but "to our knowledge, this is the first example of such a conformational mechanism proven for a biologically active small molecule that targets an RNA structure outside the bacterial ribosome," Hermann and coworkers write.

"We don't have plans to develop the benzimidazoles further because they are fairly difficult to make, not very druglike, and previously patented," Hermann tells C&EN. "We are now screening biased libraries of RNA-friendly compounds against the IRES target. The aim is to find new, synthetically more accessible and druglike classes of inhibitors that act by the same mechanism."

Hermann believes the way forward for RNA targeting is for researchers to refine the current understanding of what makes a ligand RNA-friendly while keeping druglikeness in mind. “Small molecules, not-so-small ones, and libraries thereof will have to be designed and synthesized specifically for RNA targets,” Hermann says. “This will require some compromises with regard to druglikeness, just as we see such compromises in natural antibiotics that target ribosomal RNA.” The goal is to increase “the success rate against RNA targets.”

**Another viral** RNA genome being targeted by small molecules is that of human immunodeficiency virus (HIV), the cause of AIDS. For instance, Appella and coworkers are trying to hit the transactivation response element (TAR), a component of HIV’s RNA genome that helps the virus replicate. Disabling TAR would prevent HIV replication.

Host-cell translation of HIV’s genome is boosted greatly when the HIV protein Tat binds to TAR, which has a hairpin shape. To find small molecules that bind to TAR and inhibit this interaction, Appella and coworkers synthesized and screened a library of functionalized polyamines, which are polypeptides with the backbone amides replaced by amino groups. These compounds, which they call multivalent-binding oligomers (MBOs), have nonionic side chains designed to hydrogen bond or stack with RNA bases, and their backbone amines tend to interact with the anionic RNA backbone.

The best MBOs they identified inhibit Tat-TAR binding at low micromolar levels of activity. The agents have good selectivity for TAR, enter cells easily, and have low toxicity. Structural information about the MBO-TAR complex “would be a dream come true for us so we could see how to chemically modify MBOs into more-active conformations,” Appella says.

At Clemson University, meanwhile, associate professor of bioorganic and medicinal chemistry [Dev P. Arya](#) and coworkers are also trying to develop small molecules—aminosugars, in their case—that inhibit HIV’s TAR domain. In these studies, “we are violating every possible rule of Lipinski,” Arya says, referring to Lipinski’s Rule of Five, a set of parameters for assessing a compound’s druglikeness and predicting its prospects for being orally active. Lipinski’s rule was developed for conventional drugs, which typically bind proteins, not RNAs.

Using a structure-guided approach, Arya’s group screened a library and identified modified aminosugars that bind TAR with nanomolar to sub-micromolar affinities. Some of the compounds inhibit the Tat-TAR interaction at nanomolar concentrations and protect cells from HIV infection at low toxicity levels. The compounds aren’t druglike by Lipinski’s criteria, but “our preliminary data indicate we have some promise here,” Arya said.

For inspiration in the field of small-molecule RNA targeting, Appella looks to studies in which RNA evolution experiments are used to identify RNAs with new functional properties. In those experiments, he says, “you start with enormous numbers of different RNAs and then slowly cull out, select, and amplify the ones you want.” They show that RNAs can be evolved to bind selectively to compounds such as caffeine or protein targets.

“I sense that there is selectivity in there somewhere, and we should be able to make small molecules that target RNA, too,” Appella says. “But we’re coming at it from the reverse direction”—identifying small molecules that bind RNA, not RNAs that bind small molecules—“and we don’t have the benefit of in vitro evolution, so it’s really a challenge. We have to use our ingenuity as chemists and think, ‘What are the physical properties of RNA, and how do these affect the way it likes to bind things?’ Hopefully we’ll converge on a basic idea of how to target RNA molecules.”

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